

## LC-MS compatible Separation of the Fungicide Spiroxamine

Spiroxamine is a systemic fungicide, which was brought to the market by Bayer CropScience. The substance is a mixture of diastereomers A and B again consisting of 4 enantiomers A1, A2, B1 and B2 (fig. 1).



#### Chiral Method for LC-MS

Bayer CropScience was looking for a new state-of-the-art LC-based chiral separation for spiroxamine that can be coupled to LC-MS/MS in the background of analyses of residues of spiroxamine at trace concentrations. Therefore, this meant a preference for reversed phase (RP) conditions should be investigated. Further, the aim was to perform the separation of the four isomers in less than 30 minutes. It was intended to first develop a separation application using LC-UV and then to transfer the application to LC-MS/MS.

YMC accepted the challenge and developed a new chiral application on a CHIRAL ART Amylose-SA column using water, ethanol and diethylamine.



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Fig. 1: Isomers of spiroxamine [1], diastereomer A:  $log P_{ow} = 2.79$  (at 20 °C), diastereomer B:  $log P_{ow} = 2.92$  (at 20 °C),  $pK_a$  value = 6.9.



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#### **Existing Chiral Normal Phase Separation**

Bayer CropScience had already developed a chiral method to separate these enantiomers and diasteromers respectively from each other [1]. However, at the time of the development only coated polysaccharide phases were available, which are only suitable for normal phase (NP) conditions. The resulting method was performed on a Chiralcel® OD-H column with n-heptane and isopropanol as eluents.

In a first step YMC was able to reproduce this separation on a corresponding CHIRAL ART Cellulose-C column using the same chiral selector (fig. 2).

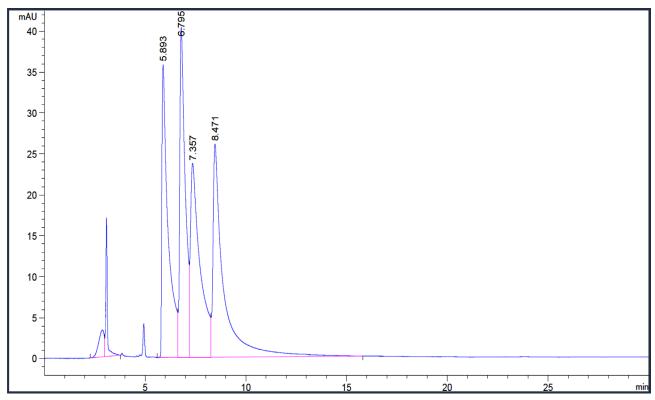


Fig. 2: Normal phase separation on CHIRAL ART Cellulose-C using the existing published method.

Column: CHIRAL ART Cellulose-C 5 µm (250 x 4.6 mm ID)

Part No.: KCN99S05-2546WT

Eluent: *n*-heptane/isopropanol (99.9/0.1)

Flow rate: 1.0 ml/min Temperature: 25 °C

Detection: UV at 210 nm Injection: 20 µL (0.1%)



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### **Chiral RP Screening**

To overcome the requirement of RP conditions, YMC tested all immobilised YMC chiral phases, namely CHIRAL ART Amylose-SA, Cellulose-SB and Cellulose-SC, with RP eluents. A sufficient separation could only be obtained on a CHIRAL ART Amylose-SA column (fig. 3). A particle size of 5  $\mu$ m and a dimension of 250 x 4.6 mm were used for each column.

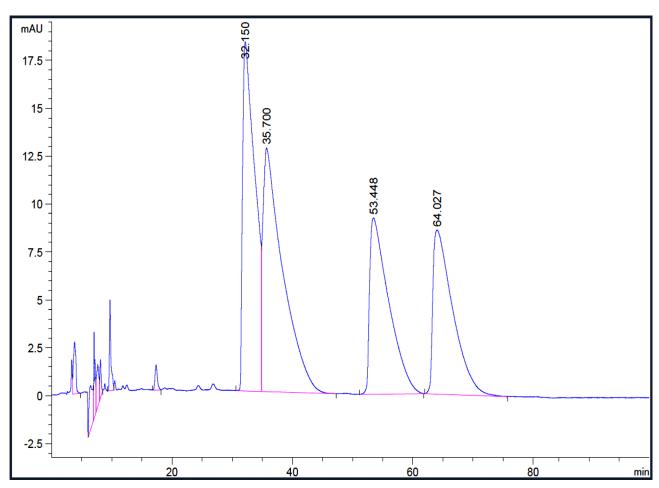


Fig. 3: Result of chiral phase screening of spiroxamine on CHIRAL ART Amylose-SA, 5 μm.



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#### Separation in less than 20 Minutes

In order to achieve the aim of a separation in less than 30 minutes, the column length and ID were reduced. In addition, 3  $\mu$ m particles were used instead of 5  $\mu$ m to increase the resolution.

The ionisation in MS detection can be improved by using an additive, but the addition of acetic or formic acid was found not useful as no retention could be observed. However, the addition of diethylamine resulted in an even better peak shape.

It was not only possible to separate all 4 of the isomers from each other, but also the separation time could be reduced to just 20 minutes. Therefore, all requirements could be fulfilled.

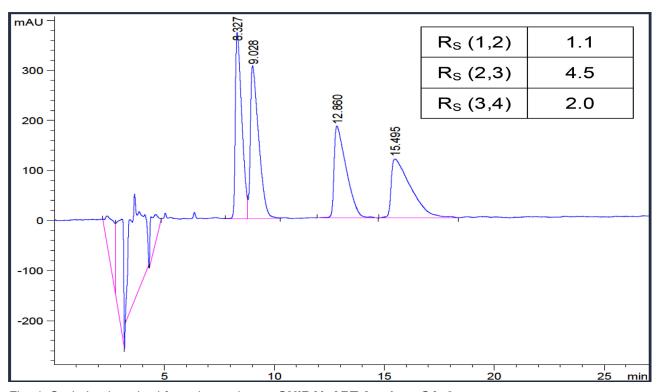


Fig. 4: Optimised method for spiroxamine on CHIRAL ART Amylose-SA, 3 μm.

Column: CHIRAL ART Amylose-SA 3 µm (150 x 3.0 mm ID)

Part No.: KSA99S03-1503WT

Eluent: water/ethanol/DEA (27.5/72.5/0.1)

Flow rate: 0.25 ml/min Temperature: 30 °C

Detection: UV at 210 nm Injection: 10 µL (10 mg/mL)



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#### Transfer from LC-UV to LC-MS/MS

At Bayer CropScience the application provided by YMC was set up on a LC-MS/MS instrument consisting of an Agilent 1290 UPLC system (binary and isocratic pump), a CTC auto-injector (Axel Semrau) and a Sciex API6500 high-end triple-quadrupole mass spectrometer.

The diethylamine used in the YMC UV-application had to be substituted by a 10 mM ammonium carbonate solution (pH 9.5). The diethylamine strongly influenced the ionisation process and "quenched" the MS-signal to about 99%.

To improve the ionisation 1% formic acid in methanol/water 50/50 was introduced post-column into the eluent flow coming from the chiral column ("change" of pH value from weak alkaline to weak acidic protonating spiroxamine; pK<sub>a</sub> 6.9).

### **Final Set-up**

Final conc. at LOQ: 1  $\mu$ g/L spiroxamine = 0.54  $\mu$ g/L A-Isomer; 0.46  $\mu$ g/L B-Isomer

= 0.27 µg/L A1 enantiomer

=  $0.27 \,\mu g/L \,A2$  enantiomer

= 0.23 µg/L B1 enantiomer

= 0.23 µg/L B2 enantiomer

HPLC Column: CHIRAL ART Amylose-SA 3 μm (150 x 3.0 mm ID)

Eluent: A/B (25/75)

A: Water/Ethanol 9/1 + 10 mM ammonium carbonate (pH ~9.5)

B: Water/Ethanol 1/9 + 10 mM ammonium carbonate

Flow: 0.3 mL/min

Inj. Volume: 1 μL

Iso-pump: post-column make-up via T-peace with 0.3 mL/min 1% formic acid in water/methanol 50/50.

MS-MS conditions: Multiple-reaction-monitoring (MRM) mode in ESI positive, MRM 298-144 for quantitation and MRM 298-100 for confirmation.



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#### **Example Chromatogram with Enantiomer A1 at LOQ in Grape Matrix Extract**

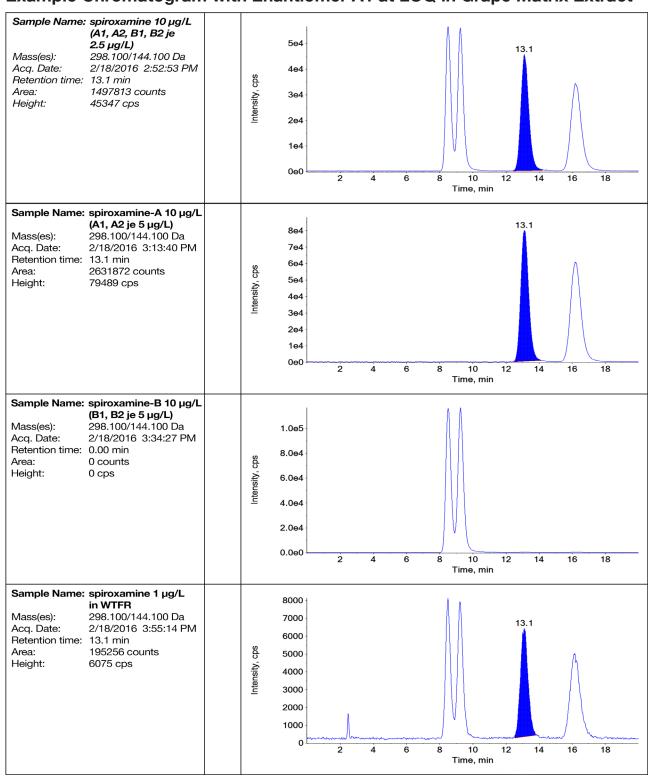


Fig. 5: Determination of enantiomer A1 in grape matrix extract, 4th repetition of each chromatogram [all chromatograms kindly provided by S. Stuke, Bayer CropScience].



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#### **Chromatographic Performance Parameter**

Study No.:

Column Type: YMC CHIRAL ART Amylose-SA 3 µm, (150 x 3.0 mm ID)

File name:

Evaluated Sample: 10 μg/L spiroxamine (mixtures of 4 enantiomers)

Col. Length [mm]	150
Col. ID [mm]	3
Pre-Col. Length [mm]	0
Pre-Col. ID [mm]	2.1
Particle Size d <sub>p</sub> [µm]	3
Col. Porosity (filling with mobile phase)	0.70
V <sub>d col</sub> [µL]	742.1
V <sub>d Pre-col</sub> [μL]	0.0
V <sub>loop</sub> [μL]	1
V <sub>capillaries</sub> [μL]	5.7
V <sub>total</sub> [µL]	748.7
flow [µL/min]	300
t <sub>d cal.</sub> [min]	2.496

theor. Plates N: 
$$N = 5.54 \left(\frac{t_R}{w_{0.5}}\right)^2$$

Peak capacity n: 
$$n = 1 + \frac{\sqrt{N_{\text{max}}}}{4} \ln(1 + k_{\text{max}})$$

Capacity Factor: should be: > 3 best: 4-10 Separation Factor  $\alpha$ :  $k'_2/k'_1$  always > 1

Peak Resolution: 0.0 = co-eluting

0.6 = 12% peak overlapping 1.0 = 2% peak overlapping 1.5 = baseline separated

Analyt = Order of Elution	t <sub>R</sub>	k'-Value	W <sub>H</sub>	W <sub>H</sub>	St. Dev. o	Theor. Plates	Plate Hight H	plates/m	Separation Factor	Peak Resolution RS <sub>n.n+1</sub>	k'- <sub>max</sub> u. N <sub>max</sub>	Peak capacity
	[min]		[min]	[S]		N	[µm]	N/m				n
spiroxamine enantiomere B1	8.50	2.41	0.3385	20.31	8.628	3494	42.9297	23294			k <sub>max</sub>	
spiroxamine enantiomere B2	9.24	2.70	0.3570	21.42	9.099	3712	40.4083	24747	1.12	1.22	5.49	29
spiroxamine enantiomere A1	13.10	4.25	0.5080	30.48	12.948	3685	40.7064	24566	1.57	4.47	N <sub>max</sub>	29
spiroxamine enantiomere A2	16.19	5.49	0.6708	40.25	17.098	3228	46.4698	21519	1.29	2.71	3712	

#### Chromatographic characteristics:

Peak capacity n 29 theor. plates/m 24747

#### Literature

[1] Etzel WA, Gau W, Krämer W, Stelzer U, Weissmüller J, Assignment of the Stereochemistry of Spiroxamine by Two-Dimensional NMR Spectroscopy and Stereoselective Chemical Synthesis, Magnetic Resonance in Chemistry, 36, 64-68 (1998).

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